

Sanger Sequencing cleanup:

Use CleanSeq beads, AMPureXP beads, or an equivalent.

1. Add 10 ul of beads to your Sanger sequencing reaction.
2. Add 80 ul of 85% EtOH.
3. Mix by pipetting up and down.
4. Put tubes on magnetic plate for 2:00 – 3:00 minutes.
5. Leaving tubes on the magnetic plate, withdraw liquid (can use vacuum).
6. Add 150 ul 85% EtOH to wash.
7. Withdraw as much liquid as possible (can use vacuum). Allow to air dry for 5:00 minutes if sending to UC Davis sequencing facility or ~ 10:00 minutes if sending to UW Madison sequencing facility.
8. For UC Davis: Remove tubes from the magnetic plate and resuspend beads in 70 ul of 0.025 mM EDTA. For UW Madison: Remove tubes from the magnetic plate and seal dried tubes. Package in bubble wrap and FedEx overnight to WI using the UCDBuy system to create a FedEx shipping label.